

See discussions, stats, and author profiles for this publication at: <https://www.researchgate.net/publication/324655536>

The Population Dynamics of a Recently Introduced Crayfish, *Cherax quadricarinatus* (von Martens, 1868), in the Sanyati Basin of Lake Kariba, Zimbabwe

Article in *African Zoology* · April 2018

DOI: 10.1080/15627020.2018.1448719

CITATIONS

0

READS

191

5 authors, including:



Lightone Marufu

University of Zimbabwe

13 PUBLICATIONS 36 CITATIONS

[SEE PROFILE](#)



Maxwell Barson

University of Zimbabwe

61 PUBLICATIONS 425 CITATIONS

[SEE PROFILE](#)



Portia Chifamba

University of Zimbabwe

11 PUBLICATIONS 85 CITATIONS

[SEE PROFILE](#)



Tamuka Nhiwatiwa

University of Zimbabwe

68 PUBLICATIONS 582 CITATIONS

[SEE PROFILE](#)

Some of the authors of this publication are also working on these related projects:



Rapid assessment of Zimbabwe's water quality using GIS and Remote Sensing technologies [View project](#)



The role of southern African bulinid snails (Mollusca: Gastropoda) in transmission of human and animal schistosomiasis and other trematode infections [View project](#)




The population dynamics of a recently introduced crayfish, *Cherax quadricarinatus* (von Martens, 1868), in the Sanyati Basin of Lake Kariba, Zimbabwe

Lightone Marufu, Maxwell Barson, Portia Chifamba, Michael Tiki & Tamuka Nhiwatiwa

To cite this article: Lightone Marufu, Maxwell Barson, Portia Chifamba, Michael Tiki & Tamuka Nhiwatiwa (2018): The population dynamics of a recently introduced crayfish, *Cherax quadricarinatus* (von Martens, 1868), in the Sanyati Basin of Lake Kariba, Zimbabwe, *African Zoology*, DOI: [10.1080/15627020.2018.1448719](https://doi.org/10.1080/15627020.2018.1448719)

To link to this article: <https://doi.org/10.1080/15627020.2018.1448719>

 Published online: 20 Apr 2018.

 Submit your article to this journal [↗](#)

 Article views: 5

 View related articles [↗](#)

 View Crossmark data [↗](#)

The population dynamics of a recently introduced crayfish, *Cherax quadricarinatus* (von Martens, 1868), in the Sanyati Basin of Lake Kariba, Zimbabwe

Lightone Marufu^{1*}, Maxwel Barson¹, Portia Chifamba¹, Michael Tiki³ and Tamuka Nhiwatiwa^{1,2}

¹ Department of Biological Sciences, University of Zimbabwe, Harare, Zimbabwe

² University of Zimbabwe Lake Kariba Research Station, Kariba, Zimbabwe

³ Zimbabwe National Parks and Wildlife Authority, Harare, Zimbabwe

* Corresponding author, email: marufulightone@gmail.com

The redclaw crayfish *Cherax quadricarinatus* is a recent introduction into Lake Kariba, Zimbabwe where it is rapidly spreading. There are, however, no studies that have investigated the population dynamics and possible ecological impacts of the crayfish in the lake. This study investigated aspects of the population biology of *C. quadricarinatus* in the lake. These included sex distribution, population growth and mortality parameters, probability of capture, recruitment and length at age of ge at maturity. Crayfish were captured monthly between January and December 2013 from 13 sites using opera house bait traps. Aspects of the species population biology were estimated using a fish stock assessment tool, FISAT II. A total of 3 205 crayfish were captured, of which 44% were male, 49% female and 7% intersex individuals. The asymptotic length ($L_{\infty} = 112.88$ mm), curvature parameter ($k = 0.72$), longevity ($t_{\max} = 4.17$ years), growth performance index ($\phi = 3.96$), total mortality ($Z = 2.06$ y⁻¹), fishing mortality ($F = 1.07$ y⁻¹), natural mortality ($M = 0.99$ y⁻¹), rate of exploitation ($E = 0.52$) and length-at-first capture ($L_c = 44.48$ mm) were estimated. Fecundity averaged 503 ± 229 eggs female⁻¹ and increased with size (weight and carapace length). Gravid females were found throughout the year but the highest proportion of females with eggs (4–10%) were found in the first quarter, whereas recruitment peaked in May and June. The smallest crayfish with eggs (48.75 mm CL) was estimated to be 1.02 years and the largest (94.82 mm CL) 2.85 years. *Cherax quadricarinatus* exhibited an *r* strategy life-history pattern due to continuous reproduction and high fecundity. In Lake Kariba, *C. quadricarinatus* is now well into the establishment stage of the Introduction–naturalisation–invasion continuum and spreading in the lake. Management options should thus aim at preventing further spread and reducing the population size and the potential negative impacts of this species.

Keywords: fecundity, mortality, recruitment, reproduction, sex distribution

Introduction

Invasive aquatic species are among the most prominent threats to biodiversity in the world (Sala et al. 2000; Clavero and García-Berthou 2005; McGeoch et al. 2010). Freshwater crayfish are keystone species that are able to affect aquatic ecosystems at varying trophic levels (Momot 1995; Lodge et al. 2000; Kats and Ferrer 2003; Chucholl 2012). They are known to disturb ecosystem functions by competing with indigenous species, destroying habitats and ultimately changing the trophic structure of the invaded environment (Lockwood et al. 2007; Davis 2009; Chucholl 2012).

Classic examples include how *Procambarus clarkii* destroyed habitats in some Spanish lakes within a few years (Rodríguez et al. 2005), the crayfish plague (*Aphanomyces astaci*) epidemic in Europe that occurred after the introduction of invasive crayfish *Pacifastacus leniusculus*, *Procambarus clarkii* and *Orconectes limosus* (Parvulescu et al. 2012) and the displacement of native fishes by *P. leniusculus* in a British lowland river (Guan and Wiles

1997). These negative impacts are some of the reasons why the introduction of crayfish has been discouraged in some parts of Africa (de Moor 2002), despite being in demand as an aquaculture species.

Cherax quadricarinatus is not indigenous to Africa (Adegboye 1983), but has been introduced for aquaculture and now occurs in South Africa (de Moor 2002; Nunes et al. 2017), Swaziland (Nunes et al. 2017), Mozambique (Nunes et al. 2017), Zambia (Thys van der Audenaerde 1994) and, more recently, in Zimbabwe (Marufu et al. 2014). *Cherax quadricarinatus* originates from northern Australia and Papua New Guinea (Austin 1996). Its introduction into Lake Kariba, which is shared between Zimbabwe and Zambia, is an example of an accidental introduction. It was initially introduced into fish farms in Livingstone, Zambia, for aquaculture purposes in 1992 (Thys van der Audenaerde 1994). The species is also believed to have escaped from another fish farm in Siavonga, Zambia from where it directly escaped into Lake Kariba (Kafue River Trust 2017). A study

by Nakayama et al. (2010) was one of the first to observe *C. quadricarinatus* in catches from the lake. The species is rapidly spreading in the lake (Marufu et al. 2014), and there is a need for management of the species.

Studies of the population biology of invasive species have been shown to be important for effective management purposes (Grandjean et al. 2000; Sakai et al. 2001; Alhassan and Armah 2011). For example, Freeman et al. (2010) recommended the use of biological control and integrated pest management options after undertaking research to better understand the population biology of the invasive signal crayfish *Pacifastacus leniusculus*. However, studies on the population dynamics of alien populations of *C. quadricarinatus* in the wild are rare (Ahyong and Yeo 2007; Vega-Villasante et al. 2015). This is in contrast to the numerous studies on its growth under aquaculture and experimental conditions (Romero 2002; Thompson et al. 2005; Barki et al. 2006). Such studies of captive crayfish populations are not easily translated to conditions in the wild (Archard and Braithwaite 2010) and give little insight into possible life-history strategies employed in the wild.

Cherax quadricarinatus is a recent introduction into Lake Kariba and there have been no comprehensive studies that assess basic biological aspects of the species, such as reproduction and growth. Only one previous study (Marufu et al. 2014) has reported on some elements of the reproductive and growth biology of *C. quadricarinatus* in the lake. That study, however, was limited to one sampling event and the results were preliminary. The aim of the present study was, therefore, to undertake a more comprehensive assessment of the reproductive and growth biology of *C. quadricarinatus* in Lake Kariba by assessing aspects of its reproductive biology and population dynamics. This information will be used to determine the stage of the introduction–naturalisation–invasion continuum of *C. quadricarinatus* in the lake (*sensu* Blackburn et al. 2011) and subsequently advise on possible management

approaches to minimise its spread and impact.

Materials and methods

Study site

The study was done in the Sanyati Basin of Lake Kariba (Figure 1). Lake Kariba is a man-made reservoir that was constructed in 1960. The lake is shared between Zimbabwe and Zambia and was constructed mainly for hydroelectric power generation. The dam height is 128 m and the lake has a maximum length of 280 km and a width of 32 km. The lake has a total surface area of 5 580 km² and an estimated volume of carrying capacity of 185 km³. The Sanyati Basin is one of the five basins of Lake Kariba. It is located in the north-eastern end of the lake. The length of the basin is 46 km with a mean width of 27 km and a total surface area of 1 223 km². The mean depth is 33 m and the total volume is 40×10^6 m³.

Crayfish capture

A total of 13 sites along the north-eastern shoreline of the Sanyati Basin were sampled monthly between January and December 2013 (Figure 1). Crayfish were trapped using the method described by Marufu et al. (2014). Six opera house activity traps (with a base length of 1 m and width of 0.5 m) were placed at each site and removed the following morning. Cooked maize meal paste was used as bait. The number of crayfish caught in each trap, as well as their carapace length (mm), mass (g) and sex were recorded. Crayfish were sexed by examining the location and type of genital pores as described by Vazquez and Lopez-Greco (2007). Individuals that could not be sex differentiated, due to possession of both male and female genital pores, were recorded as intersex individuals. Chi square analysis was used to assess if the sex ratios departed from the expected ratio (1:1) for females and males.

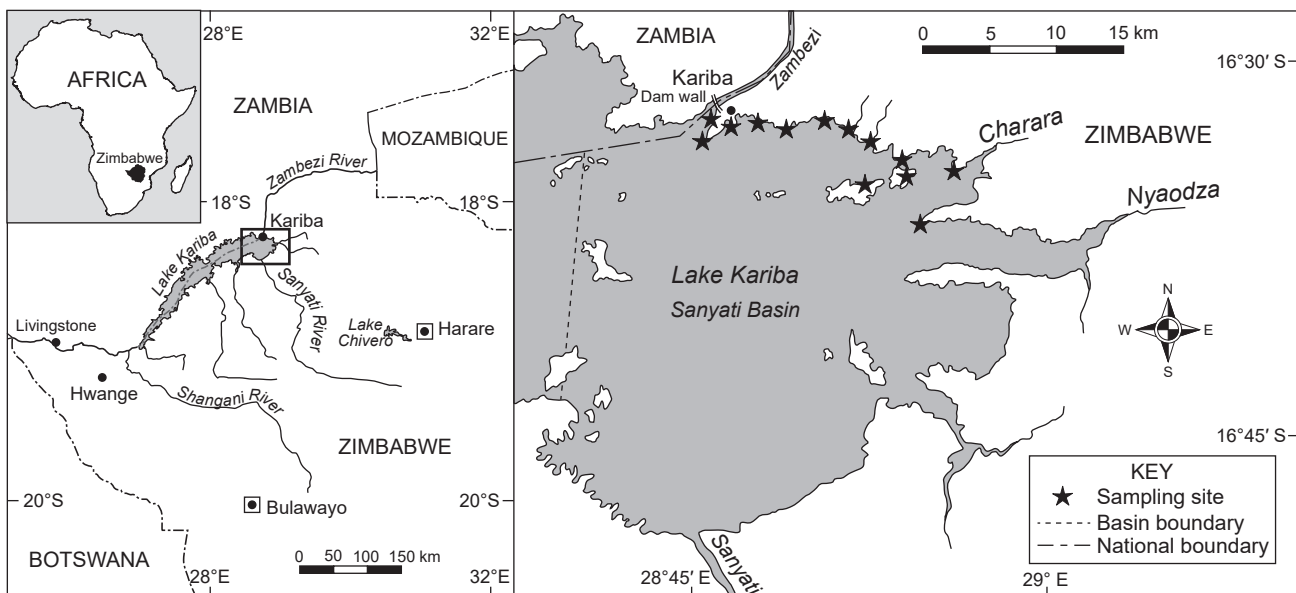


Figure 1: Map of the Sanyati Basin, Lake Kariba showing the sampling sites

Crayfish growth, longevity and mortality

Analysis of growth and mortality parameters were estimated using the FiSAT II (FAO–ICLARM Stock Assessment Tools) version 1.2.2 (2000–2005) software (Gayanilo et al. 2005). A length frequency graph was produced by classifying the carapace length frequency using 5 mm intervals and plotted using the appropriate mid-lengths.

First estimates of mortality, longevity and recruitment were also determined using FiSAT II. The instantaneous rate of annual mortality (Z) using the length-converted catch curve (Pauly 1990) and longevity (t_{max}) was estimated using the equation $t_{max} = (3/K) + t_0$, where K is the von Bertalanffy growth parameter and t_0 is the x-intercept from the fit of the von Bertalanffy growth function (Chucholl 2011, 2012).

Fecundity

A method described by Chucholl (2012) was used to determine the fecundity of *C. quadricarinatus*. All females with eggs were captured and transported to the laboratory for further analysis. All attached eggs were stripped off using forceps and counted using a counter. The weight (g) and carapace length (mm) of females with eggs were measured. In order to minimise bias due to excessive losses of eggs during transportation, only females with >300 eggs were used in further analysis. A linear model for the data was plotted and its goodness of fit value (R^2) assessed. The correlation (r) between fecundity and crayfish size (carapace length and weight) was also determined.

Results

Sex distribution

A total of 3 205 crayfish were caught between January and December 2013. Overall, females were the most abundant (49% of the total catch), whereas intersex crayfish were the least frequent (7%) (Table 1). Sex ratio was significantly different from the expected sex ratio of 1:1 ($\chi^2 = 8.77$, $df = 1$, $p < 0.05$), with proportions of the different sexes varying throughout the year (Figure 2). Between January and March, males were more active, but from April females constituted a higher proportion of the catch compared with males. The proportion of intersex individuals remained below 10% of the catch throughout the sampling period, except for the month of June in which it was 25% (Figure 2).

Growth parameters

The length frequency histograms for *C. quadricarinatus* in Lake Kariba derived from the FiSAT II analysis and the resultant von Bertalanffy growth model are shown in Figure 3. The observed maximum carapace length (L_{max})

was estimated to be 107.5 mm, whereas the predicted extreme carapace length (L_1) was 109.14 mm (95% confidence interval 104.48–113.80 mm). The curvature parameter ($K = 0.72$), asymptotic length ($L_{\infty} = 112.88$ mm) and $t_0 = -0.09$ were also derived. From the estimation of L_{∞} , K and t_0 , longevity ($t_{max} = 4.17$), growth performance index ($\phi = 3.96$) and total mortality ($Z = 2.06$) were estimated.

Fecundity

A total of 25 females with eggs were caught during the study. Length at age data were used to estimate the age of the smallest (1.02 years) and largest (2.85 years) crayfish that were caught with eggs by using the growth curve derived in Figure 3. The mean weight and carapace length of females with eggs was 71.20 ± 27.87 g and 67.08 ± 11.20 mm, respectively, with the mean number of eggs being 502.71 ± 228.78 eggs female⁻¹ (Table 2). *Cherax quadricarinatus* was generally found to be gravid throughout the year, with the highest proportion of females with eggs found in the early months of January to March and then decreasing to less than 2% from April to December (Figure 4). Fecundity increased with size (Figure 5) with regression analysis indicating a strong correlation between both length and weight with number of eggs ($R^2 = 0.62$ and $R^2 = 0.73$, respectively)

Discussion

Population dynamics studies can help to reveal important biological aspects of an alien species invasion and its progress along the introduction–naturalisation–invasion continuum (Davis 2009; Blackburn et al. 2011). The current study has provided evidence that *C. quadricarinatus* is now well into the ‘establishment stage’ on the introduction–naturalisation–invasion continuum because it has been able to survive and reproduce multiple generations since its introduction into the lake. This, in conjunction with the wide extent of occurrence of *C. quadricarinatus* in the Zambezi River Basin (see Nunes et al. 2016), shows that the species can be considered fully invasive (*sensu* Blackburn et al. 2011).

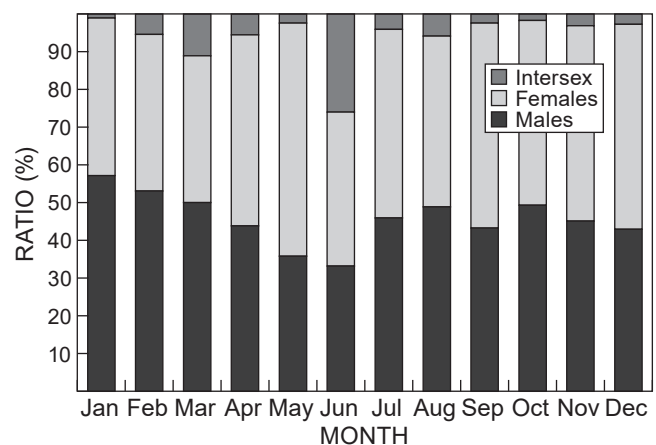


Figure 2: Changes in the sex distribution of *Cherax quadricarinatus* in the Sanyati Basin of Lake Kariba

Table 1: Distribution of sexes in *Cherax quadricarinatus* from Lake Kariba between January 2013 and December 2013

	Males	Females	Intersex	Total
Total caught	1401	1582	222	3205
Mean catch	215.53 ±	243.38 ±	34.15 ±	477.69 ±
± SD	44.47	62.09	31.12	107.07
Ratio (%)	44	49	7	100

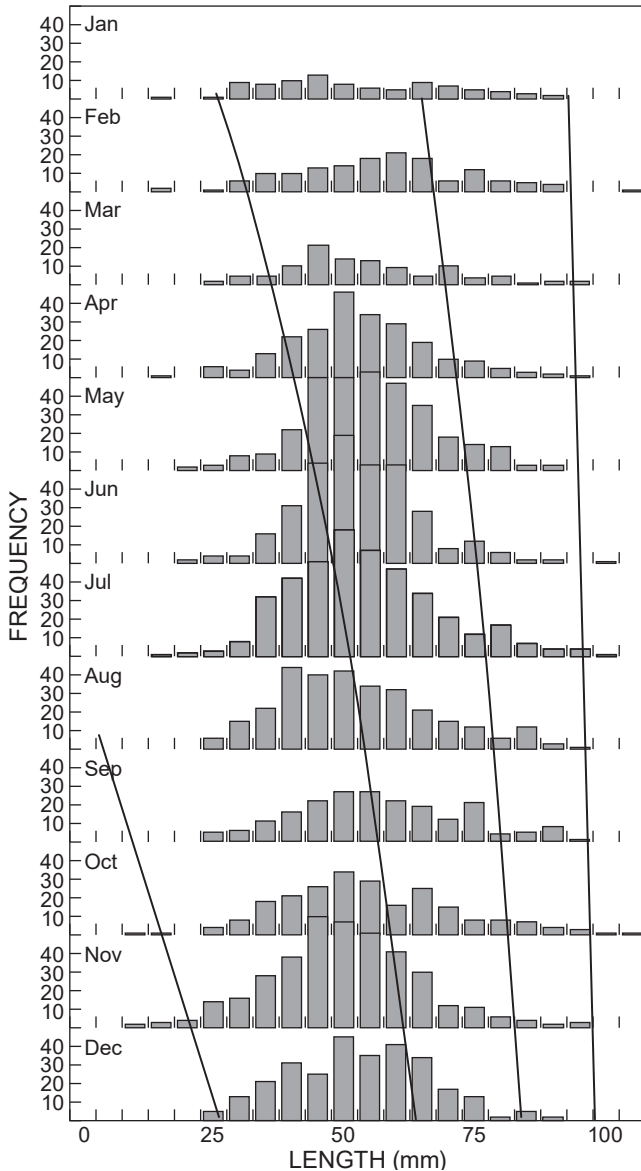


Figure 3: Estimated VBGF growth model of *Cherax quadricarinatus* caught in Lake Kariba after analysis with FiSAT II. Histograms represent the distribution of the size classes at 5 mm intervals

Table 2: Fecundity of *Cherax quadricarinatus* caught from the wild in Lake Kariba, Zimbabwe

	Mean ± SD	Minimum	Maximum
Eggs	502.71 ± 228.78	305	1 084
Weight (g)	71.20 ± 27.87	34.26	93.02
Carapace length (mm)	67.08 ± 11.20	54.09	94.82

The current study indicated that, at the onset of the year, more males were caught compared with females. The likely explanation is that most females during January–March were ovigerous, and consequently less active and therefore less likely to be caught by activity traps. Similar conclusions were made in other studies on different crayfish species also studied in the wild (Frutiger et al. 1999; Grandjean et

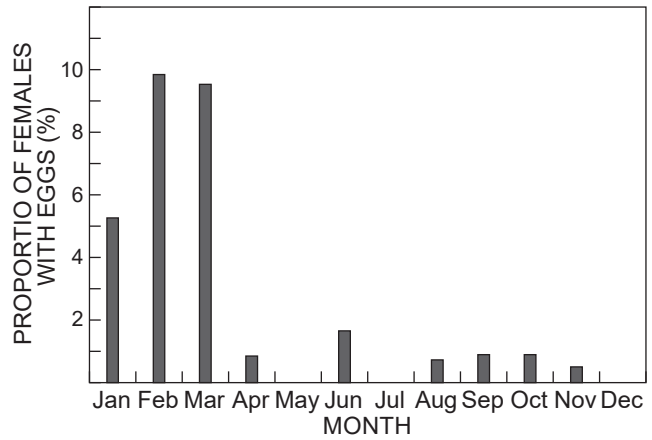


Figure 4: Proportion of *Cherax quadricarinatus* females with eggs out of total number of females caught

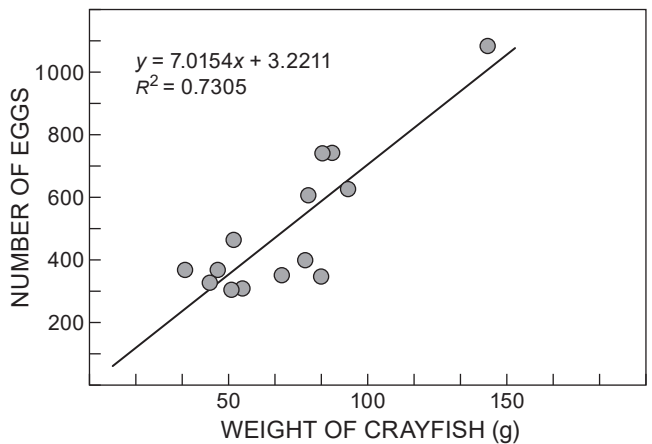


Figure 5: Linear fit between weight (g) and number of eggs of *Cherax quadricarinatus* caught in the wild of Lake Kariba, Zimbabwe

al. 2000; Dorr et al. 2006). The results also showed the occurrence of intersex individuals (with both male and female genital pores) in the population, which was not reported in previous studies on crayfish in Lake Kariba (Marufu et al. 2014; Nakayama et al. 2014). The proportion of intersex individuals in Lake Kariba (7%) was higher than the 2–4% reported by Brummet and Alon (1994) under culture conditions. Other studies (also under culture conditions) have found the proportion to be even lower at 1.2% (Sagi et al. 1996), whereas proportions as high as 17% have been reported (Medley and Rouse 1993). Intersex individuals are known to predominantly occur when culture conditions are stressful (Vazquez and Lopez Greco 2007). In addition, the occurrence of endocrine disrupting pollutants (EDPs) in crustaceans has been shown to increase the number of intersex individuals (Barki et al. 2006; Moore and Stevenson 1991). A study by Nakayama et al. (2010) found that *C. quadricarinatus* in Lake Kariba tissues contained EDPs. The EDPs present in high concentration (mg kg^{-1} dry weight) in muscle tissue included zinc (78 ± 10), copper (33 ± 8) and chromium (1.86 ± 0.98), while lead, nickel and chromium were also found in lower

amounts (Nakayama et al. 2010). This warrants further investigation.

There is some indication that *C. quadricarinatus* in Lake Kariba is following a more 'r' selected life-history strategy than in its native range. Length-at-age data revealed that *C. quadricarinatus* in Lake Kariba had a higher growth rate than for crayfish reared under controlled (aquaculture) conditions reported by Leland et al. (2015). The minimum length at maturity (estimated during the current study; 1.02 years) was similar to that reported by Rouse et al. (1991) for the same species but relatively younger when compared with a related species such as *Cherax cainii* (2–3 years) (Beatty et al. 2004) and another species *Austropotamobius torrentium* (3–4 years) (Streissl and Hodl 2002). *Cherax quadricarinatus* in Lake Kariba is producing eggs at much smaller sizes (Masser and Rouse 1997) but at higher fecundity than in its native range or under culture conditions (Sagi et al. 1997). The average number of eggs per female recorded in Lake Kariba (502) was much greater than that reported for *C. quadricarinatus* under culture conditions (321; King 1993; Sagi et al. 1997; Tropea et al. 2012). These *r* strategist characteristics are typical of an invader in a new environment. The fast growth rate, short life span, high fecundity and continuous reproduction are all indicative of an *r* strategist to ensure successful establishment in a new environment.

In conclusion, *Cherax quadricarinatus* can now be considered an established invasive species in the Zambezi basin and elsewhere (see Nunes et al. 2016, 2017). As recommended by Blackburn et al. (2011), management options at this stage of invasion should aim at containment strategies and preventing further spread. An important aspect of this will be developing monitoring programs to determine the status of this species throughout the river basin.

Acknowledgements — This study was funded by the Research Council of Zimbabwe and the University of Zimbabwe Vice Chancellor's Data Collection Fund as part of a DPhil study by Lightone T Marufu. We thank the University of Zimbabwe Lake Kariba Research Station (ULKRS) staff for their support during the study. We also appreciate the assistance received from the Zimbabwe National Parks and Wildlife Management Authority during the study.

References

- Adegboye D. 1983. On the nonexistence of an indigenous species of crayfish on the continent of Africa. *Freshwater Crayfish* 5: 564–569.
- Ahyong ST, Yeo DCJ. 2007. Feral populations of the Australian red claw crayfish (*Cherax quadricarinatus*, von Martens) in water supply catchments of Singapore. *Biological Invasions* 9: 943–946.
- Alhassan EH, Armah AK. 2011. Population Dynamics of the African River Prawn, *Macrobrachium vollenhovenii*, in Dawhenya Impoundment. *Turkish Journal of Fisheries and Aquatic Sciences* 11: 113–119.
- Archard GA, Braithwaite VA. 2010. The importance of wild populations in studies of animal temperament. *Journal of Zoology* 281: 149–160.
- Austin CM. 1996. Systematics of the freshwater crayfish genus *Cherax* Erichson (Decapoda: Parastacidae) in northern and eastern Australia: electrophoretic and morphological variation. *Australian Journal of Zoology* 44: 259–296.
- Barki A, Karplus I, Manor R, Parnes S, Aflalo ED, Sagi A. 2006. Growth of the redclaw crayfish (*Cherax quadricarinatus*) in a three-dimensional compartments system: does a neighbor matter? *Aquaculture* 252: 348–355.
- Beatty SJ, Morgan DL, Gill HS. 2004. Biology of a translocated population of the large freshwater crayfish, *Cherax cainii* (Austin and Ryan 2002) in a Western Australian river. *Crustaceana* 77: 1329–1351.
- Blackburn TM, Pysek P, Bacher S, Carlton JT, Duncan RP, Jarosik V, Wilson JRU, Richardson DM. 2011. A proposed unified framework for biological invasions. *Trends in Ecology and Evolution* 26: 333–339.
- Brummet RF, Alon NC. 1994. Polyculture of Nile tilapia (*Oreochromis niloticus*) and Australian red claw crayfish (*Cherax quadricarinatus*) in earthen ponds. *Aquaculture* 122: 47–54.
- Chucholl C. 2011. Population ecology of an alien 'warm water' crayfish (*Procambarus clarkii*) in a new cold habitat. *Knowledge in Management of Aquatic Ecosystems* 401: 29.
- Chucholl C. 2012. Understanding invasion success: life history traits and feeding habits of the alien crayfish *Orconectes immunitis* (Decapoda, Astacida, Cambaridae). *Knowledge and Management of Aquatic Ecosystems* 404: 04.
- Clavero M, Garcia-Berthou E. 2005. Invasive species are a leading cause of animal extinctions. *Trends in Ecology and Evolution* 20: 110.
- Davis MA. 2009. *Invasion biology*. Oxford: Oxford University Press.
- de Moor I. 2002. Potential impacts of alien freshwater crayfish in South Africa. *African Journal of Aquatic Science* 27: 125–139.
- Dorr AJM, La Porta G, Pedicillo G, Lorenzoni M. 2006. Biology of *Procambarus clarkia* (Girard, 1852) in Lake Trasimeno. *Bulletin Francais et de la Peche et de la Pisciculture* 380–381: 1155–1168.
- Freeman MA, Turnbull JF, Yeomans WE, Bean CW. 2010. Prospects for management strategies of invasive crayfish populations with an emphasis on biological control. *Aquatic Conservation: Marine and Freshwater Ecosystems* 20: 211–223.
- Gayanilo FC Jr, Sparre P, Pauly D. 2005. *Outils d'évaluation des stocks II (FISAT II). Version révisée. Guide d'utilisation. FAO Computerized Information Series (Fisheries) no. 8, revised version*. Rome: Food and Agriculture Organization of the United Nations.
- Grandjean F, Cornuault B, Archambault S, Brandmard M, Otrebsky G. 2000. Life history and population biology of the white clawed crayfish, *Austropotamobius pallipes*, in a brook from the Poitou-Charentes Region (France). *Bulletin Francaise de la Peche et de la Pisciculture* 356: 55–70.
- Guan RZ, Wiles PR. 1997. The home range of signal crayfish in a British lowland river. *Freshwater Forum* 8: 45–54.
- Kafue River Trust. 2017. Monitoring the spread and impact of invasive red-claw crayfish. Available at <http://www.kafue.rivertrust.org> [accessed 2 November 2017].
- Kats LB, Ferrer RP. 2003. Alien predators and amphibian declines: review of two decades of science and the transition to conservation. *Diversity and Distributions* 9: 99–110.
- King C. 1993. Potential fecundity of redclaw crayfish, *Cherax quadricarinatus* von Martens, in culture. *Aquaculture* 114: 237–241.
- Leland JC, Buchler DJ, Coughran J. 2015. Direct age determination of a subtropical freshwater crayfish (redclaw, *Cherax quadricarinatus*) using ossicular growth marks. *PLoS ONE* 10: e0134966.
- Lockwood JL, Hoopes MF, Marchetti MP. 2007. *Invasion ecology*. Oxford: Blackwell Publishing.
- Lodge DM, Taylor CA, Holdich DM, Skurdal J. 2000. Nonindigenous crayfishes threaten North American freshwater biodiversity: lessons from Europe. *Fisheries* 25(8): 7–20.

- Marufu LT, Phiri C, Nhiwatiwa T. 2014. Invasive Australian crayfish *Cherax quadricarinatus* in the Sanyati Basin of Lake Kariba: a preliminary survey. *African Journal of Aquatic Science* 39: 233–236.
- Masser MP, Rouse BB. 1997. Australian red claw crayfish. SRAC Publication no. 244. [s.l.]: Southern Regional Aquaculture Center.
- McGeoch MA, Butchrat SHM, Spear D, Marais E, Kleynhans EJ, Symes A, Chanson J, Hoffmann M. 2010. Global indicators of biological invasion: species numbers, biodiversity impact and policy responses. *Diversity and Distributions* 16: 95–108.
- Medley P, Rouse DB. 1993. Intersex Australian redclaw crayfish (*Cherax quadricarinatus*). *Journal of Shellfish Research* 12: 93–94.
- Momot WT. 1995. Redefining the role of crayfish in aquatic ecosystems. *Revision of Fisheries Sciences* 3: 33–63.
- Moore CG, Stevenson JM. 1991. The occurrence of intersexuality in harpacticoid copepods and its relationship with pollution. *Marine Pollution Bulletin* 22: 72–74.
- Nakayama SMM, Ikenaka Y, Muzandu K, Choongo K, Oroszlany B, Teraoka H, Mizuno N, Ishizuka N. 2010. Heavy metal accumulation in lake sediments, fish (*Oreochromis niloticus* and *Serranochromis thumbergi*), and crayfish (*Cherax quadricarinatus*) in Lake Itzhi-tezhi and Lake Kariba, Zambia. *Archives of Environmental Contamination and Toxicology* 59: 291–300.
- Nunes AL, Douthwaite RJ, Tyser B, Measey GJ, Weyl OLF. 2016. Invasive crayfish threaten Okavango Delta. *Frontiers in Ecology and the Environment* 14: 237–238.
- Nunes AL, Zengeya TA, Hoffman AC, Measey GJ, Weyl OLF. 2017. Distribution and establishment of the alien Australian redclaw crayfish, *Cherax quadricarinatus*, in South Africa and Swaziland. *PeerJ* 5: e3135.
- Parvulescu L, Schrimpf A, Kozubikova E, Renisino SC, Vralstad T, Petrussekand A, Schulz R. 2012. Invasive crayfish and crayfish plague on the move: first detection of the plague agent *Aphanomyces astaci* in the Romanian Danube. *Diseases of Aquatic Organisms* 98: 85–94.
- Pauly D. 1990. Length converted catch curves and the seasonal growth of fishes. *Fishbyte* 8: 33–38.
- Rodríguez CF, Bécares F, Fernández-Aláez M, Fernández-Aláez C. 2005. Loss of diversity and degradation of wetlands as a result of introducing exotic crayfish. *Biological Invasions* 7: 75–85.
- Romero X. 2002. Ups and downs of redclaw crayfish farming in Ecuador. *World Aquaculture* 33: 40–46.
- Rouse DB, Austin CM, Medley PB. 1991. Progress toward profits? Information on the Australian crayfish. *Aquaculture Magazine* 17: 46–56.
- Sagi A, Khalaila I, Barki A, Hulata G, Karplus I. 1996. Intersex red claw crayfish *Cherax quadricarinatus* (von Martens): functional males with pre-vitellogenic ovaries. *Biology Bulletin* 190: 16–23.
- Sagi A, Shoukrun R, Levy T, Barki A, Hulata G, Karplus I. 1997. Reproduction and molt in previously spawned and first-time spawning red-claw crayfish *Cherax quadricarinatus* females following eyestalk ablation during the winter reproductive-arrest period. *Aquaculture* 156: 101–111.
- Sakai AK, Allendorf FW, Holt JS, Lodge DM, Molofsky J, With KA, Baughman S, Cabin RJ, Cohen JE, Ellstrand NC, McCauley DE, O'Neil P, Parker IM, Thompson JN, Weller SG. 2001. The population biology of invasive species. *Annual Review of Ecology and Systematics* 32: 305–332.
- Sala OE, Chapin FS III, Armesto JJ, Berlow E, Bloomfield J, Dirzo R, Huber-Sanwald E, Huenneke LF, Jackson RB, Kinzig A, Leemans R, Lodge DM, Mooney HA, Oesterheld M, Poff NL, Sykes MT, Walker BH, Walker M, Wall DH. 2000. Global biodiversity scenarios for the year 2100. *Science* 287: 1770–1774.
- Streissl F, Hodl W. 2002. Growth, morphometrics, size at maturity, sexual dimorphism and condition index of *Austroptamobius torrentium* Schrank. *Hydrobiologia* 477: 201–208.
- Thompson KR, Muzinic LA, Engler LS, Webster CD. 2005. Evaluation of practical diets containing different protein levels, with or without fish meal, for juvenile Australian red claw crayfish (*Cherax quadricarinatus*). *Aquaculture* 244: 241–249.
- Thys van den Audenaerde DFE. 1994. Introduction of aquatic species into Zambian waters, and their importance for aquaculture and fisheries. ALCOM Field Document no. 24. Rome: Food and Agriculture Organization of the United Nations.
- Vazquez FJ, Lopez-Greco LS. 2007. Intersex females in the red claw crayfish, *Cherax quadricarinatus* (Decapoda: Parastacidae). *Revista de Biología Tropical* 55: 25–32.
- Vega-Villasante F, Ávalos-Aguilar JJ, Nolasco-Soria H, Vargas-Ceballos MA, Bortolini-Rosales JL, Chong-Carrillo O, Ruiz-Núñez MF, Morales-Hernández JC. 2015. Wild populations of the invasive Australian red claw crayfish *Cherax quadricarinatus* (Crustacea, Decapoda) near the northern coast of Jalisco, Mexico: a new fishing and profitable resource. *Latin American Journal of Aquatic Research* 43: 781–785.